Adula californiensis

The pea pod borer

Phylum: Mollusca

Class: Bivalvia, Pteriomorpha

Order: Mytiloida Family: Mytilidae

Taxonomy: This species was originally described under the name *Modiola californiensis* by Philippi in 1847, but was transferred to the genus *Adula* in 1857 by Adams and Adams. Additional, but uncommon, synonyms include *Adula stylina* (Huber 2010). Dall (1921) and Rocque (1953) have included *Adula* in the genus *Botula*, but Soot-Ryen (1955) differentiated the two genera. Ockelmann and Dinesen (2009) found these two genera to be distantly related, and suggested evidence for a well-defined genus *Adula* within the Mytilinae (Ockelmann and Dinesen 2009).

Description

Size: Individuals up to 40 mm in length and

10 mm in height.

Color: Shell exterior brown to black (Mytilidae, Coan and Valentich-Scott 2007), interior white, sub-nacreous, with posterior edge that is tinged with blue. Worn beaks, anteriorly, show white, and periostracum is thin, brown, and lacquer-like (Fig. 1). No chalky incrustations on shell (*Adula*, Coan and Valentich-Scott 2007).

General Morphology: Bivalve mollusks are bilaterally symmetrical with two lateral valves or shells that are hinged dorsally and surround a mantle, head, foot and viscera (see Plate 393B, Coan and Valentich-Scott 2007). Mytilids have cylindrical shells and two adductor muscles, with associated scars that are unequal in size (see Plate 395, Coan and Valentich-Scott 2007). Mytilids often use byssal threads to maintain contact with the substratum (Kozloff 1993).

Body: (see plate 17, Kozloff 1993)

Color: Interior: Exterior:

Byssus: Hairy threads which attach mussel to substrate, appear on *Adula* as a large hairy posterior patch encrusted with mud and debris (Fig. 1).

Gills:

Shell: (see Plate 403C, Coan and Valentich-Scott 2007) Valves are deep and subequal,

cylindrical, do not taper, and are thin and fragile. Dorsal and ventral margins are parallel for at least half of their length (Keep and Longstreth 1935). Umbones extend one quarter of the way from anterior end (i.e. subterminal), are not prominent, and extend higher than posterior end (Packard 1918). Shell elongate and tapers posteriorly. The posterior and anterior ends are of equal thickness (Coan and Valentich-Scott 2007).

Interior: Muscle scars very unequal in size (Mytilidae, Coan and Valentich-Scott 2007), although faint crenulations appear on the dorsal anterior margin (Fig. 2). Hinge is about one quarter of the way from anterior end.

Exterior: Surface with some radial sculpture, particularly at the anterior end, but no patches of vertical file-like striations (Fig. 1). Shells are often eroded near the beaks, anteriorly (Haderlie and Abbott 1980).

Hinge:

Eyes: Foot:

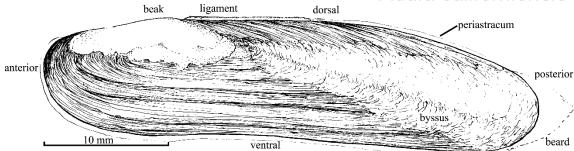
Siphons: White and fused almost to end. The incurrent siphon is with feathery oak leaf-shaped tentacles, in the illustrated specimen (Fig. 4).

Burrow:

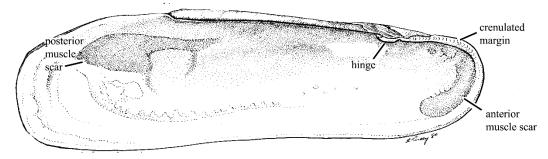
Possible Misidentifications

The family Mytilidae is characterized by two adductor muscles and associated scars that are unequal in size; the anterior scars are smaller and near the shell beak (see Plate 395, Coan and Valentich-Scott 2007). Members of this family have cylindrical shells that are dark brown or black that sometimes taper anteriorly, and the two shell valves are of similar morphology. They lack both a chondrophore (e.g., compare to *Mya arenaria*,

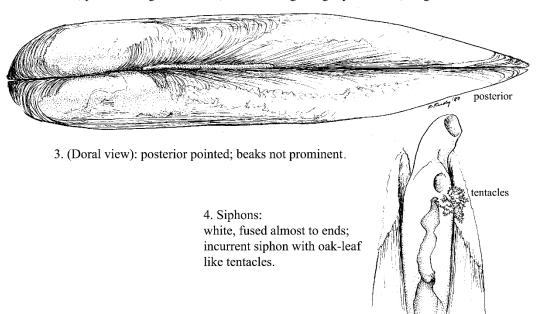
Adula californiensis



1. *Adula californiensis* x4.5: cyldindrical shells, valves subequal; dorsal and ventral margins roughly parallel; smooth periostracum; posterior slope hairy; worn beaks 1/4 of way from rounded anterior; ligament external; radial sculpture.



2. Left valve, interior: posterior muscle scar much larger than anterior scar; color white; sub-nacreous, posterior tinged with blue; anterior margin slightly crenulate; hinge without teeth.



this guide) and dorsal margin ears, and the shell is not cemented to the substratum (Coan and Valentich-Scott 2007). There are nearly 20 local species in the family Mytilidae *Modiolus* (six species) and *Mytilus* (one to four species). Other genera are possibly present in the area, but are not included in current keys including *Crenella* (*C. decussata*), *Gregariella* (*G. coarctata*), *Solamen* (*S. columbianum*) (Coan and Valentich-Scott 2007).

Genera included in the current key can be differentiated as follows: Mytilus species (see M. trossulus, this guide) have shells that with beaks at the terminal portion of the anterior end and lack internal septa, while Septifer species possess an internal septum at their anterior end. All other genera have shells with beaks that are anterior, but not terminal. Of those, Lithophaga plumula individuals are with cylindrical shells and a posterodorsal slope that is rough and with chalky encrustations, while Adula species have a posterodorsal slope without chalky encrustations and, instead, have a thick mat and are sometimes covered with mud or debris. The genera Geukensia, Modiolus, and Musculista have shells that are not cylindrical, G. demissa shells have prominent ribs externally and are dark brown or black in color. Modiolus and Musculista species have shells without external ribbing and members of the former genus have periostracial hairs while Musculista do not.

The genus closest to Adula is Lithophaga (the "date shell"), a boring mussel with cylindrical shell and roughly parallel margins. It lacks the hairy posterior of Adula, and bores in hard rock and it has peculiar feather-like wrinkling on the posterior of the shell. There are three local species of Adula including A. gruneri, A. diegensis, and A. californiensis. Adula gruneri (=A. falcata, Lithodomus falcatus, Ockelmann and Dinesen 2009) and A. californiensis both bore into shale, while A. diegensis is free-living. Adula gruneri can be recognized from the other two by the presence of persiostracum that is with irregular striae. Adula californiensis and A. diegensis can also be differentiated as the former species has an elongate shell, while

comprising the genera Septifer (S. bifurcatus), Lithophaga (L. plumula), Geukensia (G. demissa), Musculista (M. senhousia),

A. diegensis has a stout shell. Adula diegensis also has a dorsal margin that flares and sparse periostracal mat on the posterior slope, neither are present in A. californiensis. Adula diegensis is a small species (< 19 mm in length) that occurs as far north as San Francisco, California, on mudflats and pilings with other mussels (e.g., see Mytilus trossulus, this guide). It is polished and dark blue interiorly. Adula gruneri, the hooked pea-pod shell, bores deep into hard rock as well as into clay. It has wrinkled periostracum, not a smooth one, as well as vertical striae to assist in boring. Its shell is more angular and proportionally longer than the more cylindrical than A. californiensis, and tapers posteriorly. The beaks in A. gruneri are situated at about the anterior eighth of the shell length, and are strongly involute (closely wound). Its northern limit is probably Coos Bay and is known from southern Oregon to Baja California, as is A. diegensis (Kozloff 1993). Adula gruneri is the largest of the three Adula species, at up to 80 mm in length, while A. diegensis is generally smaller (~20 mm) than A. californiensis (Kozloff 1993). (For in depth differentiation between Adula species, see Ockelmann and Dinesen 2009.)

Ecological Information

Range: Type region in northwestern Pacific (Bernard 1983). Known range includes Vancouver Island, British Columbia, Canada, Alaska, and San Diego, California (Kozloff 1993).

Local distribution: Local distribution at sites in Coos Bay and Pigeon Point, also Yaquina and Boiler Bays (see Fig. 1, Lough and Gonor 1971).

Habitat: Burrows into soft, muddy shale or is, occasionally, free-living (Coan and Valentish-Scott 2007). In Coos Bay, they can be found in in old pholad (fam. Pholadidae) burrows. Distribution is limited by appropriate burrowing substrate (Lough and Gonor 1971). Salinity: Collected at salinities of 30, usually in the lower reaches of the bay where salinity

is relatively high and constant. Estuarine and marine habitats (Lough and Gonor 1971). **Temperature:** Individuals occur in temperate waters.

Tidal Level: Intertidal to sublittoral (Coan and Valentich-Scott 2007). Individuals collected from up to 10 meter depths (Newport, OR, Lough and Gonor 1971). **Associates:** Associates include the terebellid polychaete *Thelepus*, the pholad *Penitella*, and the brachyuran crab, *Cancer oregonensis*. In addition, *A. californiensis* hosts several ciliated protozoans including *Raabella* (=*Hypocomides*) *botulae*, *R. parva* and *Insignicoma venusta* (Coan and Valentich-Scott 2007).

Abundance: not common

Life-History Information

Reproduction: Dioeceous, and freespawning. Individuals are ripe from June to October (Oregon, Kozloff 1974; Lough and Gonor 1971). Oocytes are 70–80 μm in diameter and are pink to orange in color and yolky (Lough and Gonor 1971). For sperm morphology of members of the Mytilidae, including the conger, A. falcatoides, see Kafanov and Drozdov 1998.

Larva: Bivalve development generally proceeds from external fertilization via broadcast spawning through a ciliated trochophore stage to a veliger larva. Bivalve veligers are characterized by a ciliated velum that is used for swimming, feeding and respiration. The veliger larva is also found in many gastropod larvae, but the larvae in the two groups can be recognized by shell morphology (i.e., snail-like versus clam-like). In bivalves, the initial shelled-larva is called a D-stage or straight-hinge veliger due to the "D" shaped shell. This initial shell is called a prodissoconch I and is followed by a prodissoconch II, or shell that is subsequently added to the initial shell zone. Finally, shell secreted following metamorphosis is simply referred to as the dissoconch (see Fig. 2, Brink 2001). Once the larva develops a foot, usually just before metamorphosis and loss of the velum, it is called a pediveliger (see Fig. 1, Kabat and O'Foighil 1987; Brink 2001). (For generalized life cycle see Fig. 1, Brink 2001). Development in A. californiensis was described by Lough and Gonor (see Fig. 2,

1971) and proceeds with polar lobe formation within the first hour, which is resorbed by 1.5 hours, first and second cleavages at 1.5 and 2.5 hours post fertilization, respectively, trochophore larvae develop at 15 hours, the shell gland forms at at 31 hours, and a bivalve shell, that surrounds individuals entirely, is complete at 72 hours (at 15°C, Lough and Gonor 1971). Veliger larvae are freeswimming (Brink 2001) and are relatively tolerant of low salinity but not high temperatures (Kabat and O'Foighil 1987). While trochophore larvae tend to swim in all directions, veligers swim vertically and not horizontally (Lough and Gonor 1971; Haderlie and Abbott 1980). Settlement and metamorphosis occur after three days. Larval size is approximately 108 µm at 3 days, 117 µm at 15 days and 119 µm at 25 days post fertilization (Lough and Gonor 1971). Development increases with temperature and slows with decreasing salinity, with lowest salinity for development 26.3 (Lough and Gonor 1971). Young larvae (3 days old) are more sensitive to a reduction in salinity than older larvae (15 days old or more), but the opposite trend is seen for temperature tolerance (older larvae have a narrower temperature tolerance) (Lough and Gonor 1973a, b). Ideal temperature and salinity for development is 10–15°C and salinity 31–33 (Lough and Gonor 1973a).

Juvenile: Juveniles morphology flares posteriorly rather than being cylindrical (i.e. modioliform).

Longevity:

Growth Rate:

Food: A suspension feeder. For feeding mechanisms and concentration of particles using currents produced by cilia in the congener, *A. gruneri*, see Fankboner 1971.

Predators:

Behavior: This species is probably more of a nestler than a burrower.

Bibliography

 BERNARD, F. R. 1983. Catalogue of the living bivalvia of the eastern Pacific Ocean: Bering Strait to Cape Horn. Ottawa: Dept. of Fisheries and Oceans, Ottawa.

- BRINK, L. A. 2001. Mollusca: Bivalvia, p. 129-149. *In:* Identification guide to larval marine invertebrates of the Pacific Northwest. A. Shanks (ed.). Oregon State University Press, Corvallis, OR.
- 3. COAN, E. V., and P. VALENTICH-SCOTT. 2007. Bivalvia, p. 807-859. *In:* The Light and Smith manual: intertidal invertebrates from central California to Oregon. J. T. Carlton (ed.). University of California Press, Berkeley, CA.
- DALL, W. H. 1921. Summary of the marine shellbearing mollusks of the northwest coast of America, from San Diego, California to the Polar Sea. Bulletin of the United States Natural Museum. 112:1-217.
- 5. FANKBONER, P. V. 1971. Ciliary currents associated with feeding, digestion, and sediment removal in *Adula* (Botula) *falcata* Gould 1851. Biological Bulletin. 140:28-45.
- HADERLIE, E. C., and D. P. ABBOTT. 1980. Bivalvia: the clams and allies, p. 355-410. *In:* Intertidal invertebrates of California. R. H. Morris, D. P. Abbott, and E. C. Haderlie (eds.). Stanford University Press, California.
- 7. HUBER, M. 2010. Compendium of bivalves: a full-color guide to 3,300 of the world's marine bivalves. A status of bivalvia after 250 years of research. Conch Books, Hackenheim.
- KABAT, A. R., and D. O'FOIGHIL. 1987. Phylum Mollusca, Class Bivalvia, p. 309-353. *In:* Reproduction and development of marine invertebrates of the Northern Pacific Coast. M. F. Strathmann (ed.). University of Washington Press, Seattle, WA.
- KEEN, A. M., and E. COAN. 1974.
 Marine molluscan genera of western North America: an illustrated key. Stanford University Press, Stanford, CA.
- KEEP, J., and J. LONGSTRETH.
 1935. West coast shells: a description in familiar terms of the principal marine, fresh-water, and land mollusks of the United States, British Columbia, and Alaska, found west of the Sierra.

- Stanford University Press, Stanford, CA
- 11. KOZLOFF, E. N. 1974. Keys to the marine invertebrates of Puget Sound, the San Juan Archipelago, and adjacent regions. University of Washington Press, Seattle.
- 12. —. 1993. Seashore life of the northern Pacific coast: an illustrated guide to northern California, Oregon, Washington, and British Columbia. University of Washington Press, Seattle.
- 13. LA ROCQUE, A. L. 1953. Catalogue of the recent Mollusca of Canada. E. Cloutier, Queen's Printer, Ottawa.
- 14. LOUGH, R. G., and J. J. GONOR. 1971. Early embryonic stages of *Adula califoriensis* and the effect of temperature and salinity on developmental rate. Marine Biology. 8:118-125.
- 15. —. 1973a. A response-surface approach to the combined effects of temperature and salinity on the larval development of *Adula californiensis* (pelecypoda:mytilidae) I. survival and growth of three and fifteen-day old larvae. Marine Biology. 22:241-250.
- 16. —. 1973b. A response-surface approach to the combined effects of temperature and salinity on the larval development of *Adula californiensis* (Pelecypoda:Mytilidae) II. long-term larval survival and growth. 22:295-305.
- 17. OCKELMANN, K. W., and G. E. DINESEN. 2009. Systematic relationship of the genus *Adula* and its descent from a *Mytilus*-like ancestor (Bivalvia, Mytilidae, Mytilinae). Steenstrupia. 30:141-152.
- 18. PACKARD, E. L. 1918. Molluscan fauna from San Francisco Bay. Zoology. 14:199-452.
- RICKETTS, E. F., and J. CALVIN.
 1952. Between Pacific tides: an account of the habits and habitats of some five hundred of the common, conspicuous seashore invertebrates of the Pacific Coast between Sitka, Alaska, and Northern Mexico.

 Stanford, Stanford University Press, Stanford.

- 20. —. 1971. Between Pacific tides. Stanford University Press, Stanford, California.
- 21. SOOT-RYEN, J. 1955. A report on the family Mytilidae (Pelecypoda). Allan Hancock Pacific Expedition. 20:1-174.
- 22. WILSON, B. R. 2006. A new generic name for a burrowing mytilid (Mollusca: Bivalvia: Mytilidae). Molluscan Research. 26:89-97.